

Synthesis and QSAR of Novel Ketoprofen–Chalcone-Amide Hybrides as Acetylcholinesterase (AChE) Inhibitors for Possible Treatment of Alzheimer Disease

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Abstract—A new series of the anti-inflammatory drug ketoprofen derivatives bearing aryl chalcone-amide congeners were synthesized. The structures of the synthesized compounds were identified by the ¹H NMR, ¹³C NMR, and EIMS spectroscopic methods. The inhibitory activity of the synthesized compounds on cholinesterase enzymes was investigated. Biological results revealed that five compounds displayed moderate activities against acetylcholinesterase (AChE) with IC₅₀ values below 10 μM. Among the tested compounds, (BTPPh) was found to be the most potent against AChE (IC₅₀ 0.98 ± 0.02 μM), while the chalcone-amide analogues (MeOPh), (HydPh), (FPh), and (ChPh) exhibited moderate activities with IC₅₀ values ranged between 5.19–9.61 μM. Molecular docking study showed that compound (BTPPh) could combine with the active site of acetylcholinesterase by the π–π between the ketoprofen phenyl groups is embedded in a cavity surrounded by two aromatic residues of Tyr334 and Trp279. The present results strongly suggest that the *para*-position of the D-ring should be a preferred modification site for further structural optimization design. Thus, compound (BTPPh) emerged as a promising lead for the development of new acetylcholinesterase inhibitor agent. The preliminary quantum structure-activity relationship (QSAR) among the newly synthesized congeners was obtained by Genetic Function Approximation (GFA).

Keywords: alzheimer disease, acetylcholinesterase (AChE) inhibitors, chalcones, ketoprofen, molecular docking study

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INTRODUCTION

Alzheimer's disease (AD) is a multifactorial neurodegenerative dementia, characterized by deterioration of memory and cognition in elder patients [1, 2]. The main cause of the loss of cognitive functions in AD patients is a continuous decline of the cholinergic neurotransmission in cortical and other regions of the human brain [3]. Cholinergic neurotransmission is mediated by the neurotransmitter acetylcholine (ACh), which is released and carried out the effect followed by rapidly hydrolysis via acetylcholinesterase

(AChE) to acetate and choline [4, 5]. One of successful therapeutic strategies for treatment of AD is based on the cholinergic hypothesis [6]. This hypothesis assumes that in AD, the level of acetylcholine (ACh) is reduced due to the loss of the cholinergic neurons and decreased synthesis of ACh [3, 7]. The major therapeutic target in the AD treatment strategies is the inhibition of brain AChE to increase the ACh level in brain, causing increase in the cholinergic neurotransmission in AD patients [8, 9]. At present, there are four FDA-approved AChE inhibitor-type drugs, tacrine (**I**), donepezil (**II**), galantamine (**III**) and rivastigmine (**IV**) (Fig. 1) for treatment of cognitive dysfunction and memory loss associated with mild-to-moderate AD [10]. Recently, various potential analogues have been synthesized and evaluated for their activity against AChE such as indoles [11, 12], pyrroles [13, 14] chalcones [15–17], thiophene analogues [18], coumarin derivatives [19–21] and other potential

Abbreviations: BTPPh, 2-(3-benzoylphenyl)-*N*-(4-(3-(4-tolyl)acryloyl)phenyl)propanamide; MePPh, 2-(3-benzoylphenyl)-*N*-(4-(3-(4-methoxyphenyl)acryloyl)phenyl)propanamide; HydPh, 2-(3-benzoylphenyl)-*N*-(4-(3-(4-hydroxyphenyl)acryloyl)phenyl)propanamide; FPh, 2-(3-benzoylphenyl)-*N*-(4-(3-(4-fluorophenyl)acryloyl)phenyl)propanamide; ChPh, 2-(3-benzoylphenyl)-*N*-(4-(3-(4-chlorophenyl)acryloyl)phenyl)propanamide.

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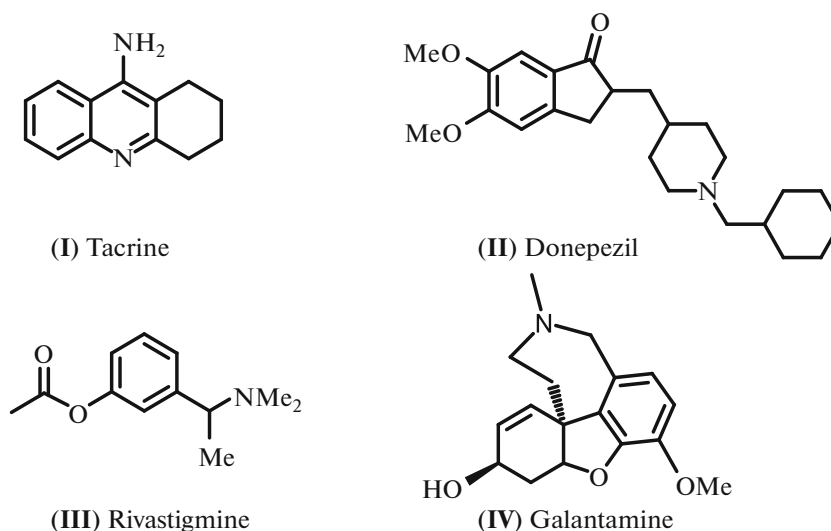


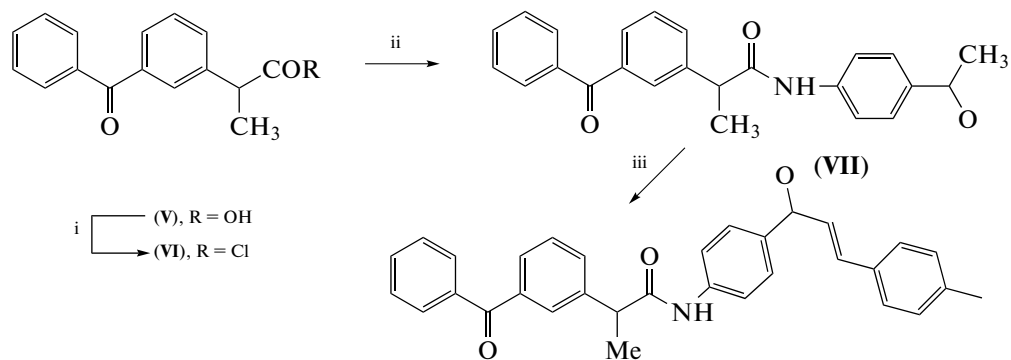
Fig. 1. Some of the acetylcholinesterase (AChE) inhibitor-type drugs.

compounds [22]. Design strategy and advantages of AChE inhibitors have been extensively reviewed [23–25].

Chalcones exhibited diverse biological activities [26], but few investigations were carried out by pharmaceutical researchers for examining their AChE inhibition activity. Based on the design experiences from existing AChE inhibitors, we synthesized a series of new chalcone-amide derivatives derived ketoprofen with evaluation of their AChE inhibition activity. Furthermore, molecular docking study was carried out for compound as well as donepezil for comparison purposes and to study the binding mode and selectivity of this analogue against AChE.

RESULTS AND DISCUSSION

The anti-inflammatory drug ketoprofen (2-(3-benzoylphenyl)propanoic acid, **(V)**) has been selected as a key intermediate for the synthesis of new substituted chalcone-amide derived ketoprofen derivatives, aiming at the evaluation of their AChE inhibition activity, *via* three steps. Thus, treatment of **(V)** with PCl_5 at 50°C for 30 min. gave the acyl chloride analogue **(VI)**, followed by simultaneous addition of 4-aminoacetophenone to the amide derivative. Treatment of the crude product with various substituted aryl benzaldehydes in the presence of NaOH afforded, after chromatographic purification, the chalcone-amide analogues **(VIII–XVII)** in 60–73% yield (Scheme 1).



	(VII)	(IX)	(X)	(XI)	(XII)	(XIII)	(XIV)	(XV)	(XVI)	(XVII)
X	H	Me	OMe	OH	NH ₂	NMe ₂	NO ₂	F	Cl	Br

Scheme 1. Synthesis of some novel ketoprofen-chalcone-amide hybrids **(VIII)–(XVII)**.

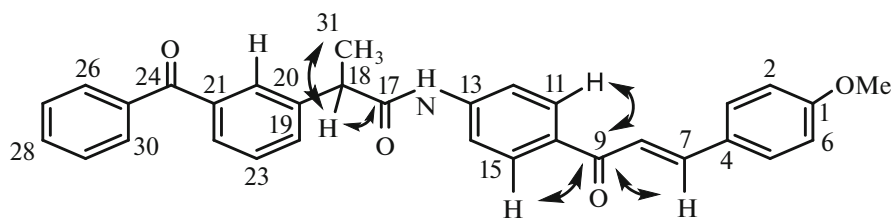


Fig. 2. $J_{C,H}$ correlations in the HMBC NMR spectrum of compound (XII).

The structures of compounds (VIII)–(XVII) were confirmed by the IR, ^1H , and ^{13}C NMR spectra. The IR spectra of (VIII)–(XVII) were characterized by the appearance of the absorption band at $1631\text{--}1705\text{ cm}^{-1}$, attributed to the (C=O) stretching, while the strong absorption bands in the range $1585\text{--}1600\text{ cm}^{-1}$, were assigned to (C=C) stretching group. In addition, the spectra showed weak absorption bands appeared in the range $3036\text{--}3053\text{ cm}^{-1}$, were due to the (CH) stretching. The medium absorption bands at the regions 3455 and $3228\text{--}3335\text{ cm}^{-1}$, were assigned to the OH and NH_2 groups of compounds (XI) and (XII). In the ^1H NMR spectra of compounds (VIII)–(XVII), the olefinic proton of chalcone moiety H-7 appeared as a broad singlet at the regions δ 8.54–7.90 ppm, while H-8 of the same group overlapped with the aromatic protons at the regions δ 8.10–7.88 and 7.38–7.25 ppm. H-18 appeared as a broad singlet at the regions δ 3.46–3.31 ppm, whereas the singlet at the regions δ 1.39–1.30 ppm assigned to Me-31.

The aliphatic protons were fully analysed (c.f. Experimental section). In the ^{13}C NMR spectra of compounds (VIII)–(XVII), the carbonyl carbon atoms of the chalcone scaffold $\text{C}_9=\text{O}$ resonated at the regions δ 172.3–170.0 ppm, whereas the carbonyl carbon atoms $\text{C}_{17}=\text{O}$ and $\text{C}_{24}=\text{O}$ were appeared at δ 191.9–189.7 ppm. C-7 and C-13 resonated together at the regions δ 146.6–143.9 ppm, while C-21 appeared at the regions δ 139.0–136.5 ppm. The resonances at the regions δ 137.1–133.3 assigned to carbon atoms C-19 and C-25 as overlapped signals, whereas C-8 appeared at the regions δ 126.7–121.9 ppm. The signals at the regions δ 44.0–40.1 ppm attributed to the carbon atom C-18. Carbon atom C-F of compound (XV) appeared as a doublet at δ 157.6 ($J_{C,F} = 247$ Hz). The other aromatic and aliphatic substituents carbon atoms were fully analysed (c.f. Experimental section). Compound (XII) was selected for further NMR experiments. From a gradient HMBC [27] NMR spectrum, the carbonyl carbon atom of the chalcone moiety $\text{C}_9=\text{O}$ at δ 189.7 ppm showed three $^3J_{C,H}$ couplings with H-7 proton at δ 7.96 ppm as well as with H-11 together with H-15 at δ 7.88 ppm, respectively. Further, H-18 at δ 3.44 ppm exhibited two $^2J_{C,H}$ couplings: one with carbonyl carbon atom $\text{C}_{17}=\text{O}$ at δ 171.5 ppm and the other

coupling with methyl carbon atom Me-31 at δ 19.8 ppm, respectively (Fig. 2).

The mass spectra of the prepared compounds showed the correct molecular ions suggested by their molecular formulas.

In vitro AChE inhibition activity. Acetylcholinesterase (AChE) is an enzyme that catalyzes the breakdown of acetylcholine and of some other choline esters that function as neurotransmitters. AChE is found at mainly neuromuscular junctions and in chemical synapses of the cholinergic type, where its activity serves to terminate synaptic transmission. An acetylcholinesterase inhibitor (often abbreviated AChEI) or anticholinesterase is a chemical or a drug that inhibits the acetylcholinesterase enzyme from breaking down acetylcholine, thereby increasing both the level and duration of action of the neurotransmitter acetylcholine. In the present work, compounds (VIII)–(XVII) were screened for their inhibitory potential against acetylcholinesterase, following the Ellman method [28] and using galantamine as the positive control. The results are presented in Table 1 with varying degrees of inhibition pattern, where the IC_{50} values of compounds are in the micromolar range. The results indicated that all compounds exhibited good to moderate inhibitory activities against AChE. Interestingly, compound (IX), exhibited the strongest AChE inhibitory activity ($\text{IC}_{50} = 0.98\ \mu\text{M}$) of the series, whereas compounds (X), (XI), (XIII), (XV), and (XVI) showed a moderate activity against AChE with IC_{50} values of 5.19, 12.19, 8.18, 9.61, and 6.61 μM , respectively. From these results it is evident that the methyl group as a substituent on the phenyl-chalcone scaffold has a vital role in the inhibition of acetylcholinesterase. The replacement of methyl to methoxy group, as in the case of product (X), will make the compound activity approximately 5-fold less effective against AChE. The least activity was observed in compound (XIV), which has the 4-nitro substituent on the phenyl-chalcone moiety with IC_{50} of 58.94 μM against AChE, while the 4-chloro or 4-fluoro substituents on the same moiety, as electron-withdrawing groups, led to marked improvement in the activity against AChE ($\text{IC}_{50} = 6.61$ and 9.61 μM , respectively). Further, it was found that the 4-bromo substituted compound (XVII) is less active ($\text{IC}_{50} = 21.27\ \mu\text{M}$) than the chloro and fluoro derivatives. However, compound (XI) with 4-hydroxyl sub-

Table 1. In vitro IC₅₀ values of ketoprofen–chalcone–amide hybrids (VIII)–(XVII) on AChE activity

Comp.	(IC ₅₀ , μM) ^a
(VIII)	19.24 ± 0.4
(IX)	0.98 ± 0.02
(X)	5.19 ± 0.41
(XI)	12.19 ± 0.97
(XII)	22.9 ± 1.26
(XIII)	8.18 ± 0.1
(XIV)	58.9 ± 4.6
(XV)	9.61 ± 0.6
(XVI)	6.61 ± 0.4
(XVII)	21.27 ± 1.1
Galantamine ^b	0.5 ± 0.01

^a Data are expressed as means ± standard deviation (SD) of at least three independent experiments. ^b Standard inhibitor of acetylcholinesterase (AChE).

stituent exhibited a moderate activity against AChE. The structure–activity relationship study revealed that the activity is greatly dependent on the type and nature of the substituents. Figure S1 (Supplementary Material) shows the inhibition (%) of the acetylcholinesterase versus the concentrations of compounds (VIII–XVII).

Molecular modelling analysis. Docking studies were performed in order to gain more insight into the binding mode of the most active compounds of the chalcone amide derivatives derived ketoprofen to the AChE enzyme. The structures of the studied ligands were prepared with the MOE interface and minimized using the default procedure in MOE (Molecular Operating Environment (MOE), 2015.10 [29]; Chemical Computing Group Inc.: Montreal, QC, Canada) with the Amber10:EHT force field. The crystal structure of the receptor with ID 1EVE was retrieved as a PDB file from the PDB base. The co-crystallized waters and the NAG ligands were stripped out before docking. The protein structure was corrected and hydrogen atoms were added and the protein–native ligand structure was then energy minimized using the Amber10:EHT force field. In docking studies were used the Stochastic search method and London dG scoring function, which is implemented in MOE. Root Mean Squared Distance (RMSD) corresponding atoms in an initial conformer and a docked pose was used to evaluate the quality of self-docking and cross-docking studies. The satisfactory values for RMSD were those lower than or equal to 2 Å.

In our search for new lead derivatives as acetylcholinesterase (AChE) inhibitor, we have selected com-

pound (IX) for molecular docking study. Thus, molecular docking of conjugate (IX) into the three-dimensional AChE (PDB ID: 1EVE) was performed using the MEO program [29]. The prospective ligands were ranked according to the highest energy of the best conformers. The calculated binding energy scores for compound (IX) is $-9.51 \text{ kcal mol}^{-1}$, indicating selectivity binding of this analogue to the active site of the protein receptor pocket (1EVE). The chalcone moiety of ketoprofen backbone oriented in an appropriate position for its binding with the amino acids of AChE via the π – π stacking interaction with Trp279 and Tyr334. Additionally, non-bonded amino acid residues surrounded (IX) were observed. In addition, non-bonded amino acid residues such as Gln74, Tyr84, Tyr70, Tyr130, Leu127, Tyr116, Glu199, Asp72, Ser122, Phe331, Phe330 and Ser386 of the receptor surrounded the compound (IX) were observed, which would enhance its inhibitory potency.

Quantitative structure–activity relationship (QSAR).

QSAR plays a crucial role in drug development as it analyzes the properties of the drug. It is a mathematical model that links the structural features of the compounds (i.e. molecular descriptors) to their quantity showing specific biological or chemical activity [30]. Therefore, it is important in QSAR study to establish the relationship relationship between IC₅₀ and numerous parameters by regression models. In this study, we investigated the structural features and conformational behaviour and the optimized geometries of building blocks of chalconyl steroids and their analogues, at the PWC/DNP level of theory using the software Dmol [31] in Materials Studio package. The QSAR study was done with the Materials Studio package using genetic function approximation (GFA) technique [32]. The quantum chemical indices E_{HOMO} , E_{LUMO} , and molecular flexibility were calculated with VAMP software. This will serve as a basis for future theoretical and experimental work on more complex aromatic steroid structures related to their biological activity. A set data of ten ketoprofen carrying chalcone–amide derivatives with selected descriptors has been chosen to study their QSAR using GFA method to generate a linear model with two varied equations (1) and (2) obtained for calculating predictive AChE inhibition activity as shown in Tables 2 and 3.

$$\begin{aligned}
 \text{pIC}_{50} = & 3.040\text{ramp}(A \log P - 11.152) \\
 & - 1.594\text{ramp}(A \log P - 11.525) \\
 & + 264.212\text{ramp}(E_{\text{LUMO}} + 0.621) \\
 & - 1107.896\text{ramp}(E_{\text{LUMO}} + 0.670)^2 + 8.289, \quad (1) \\
 R^2 = & 0.999; \text{adjusted } R^2 = 0.999 \\
 & \text{cross validated } R^2 = 0.998; \\
 & \text{significant regression} = \text{yes}; \\
 & \text{critical SOR } F \text{ value} = 2478.
 \end{aligned}$$

$$\begin{aligned}
 & \text{pIC}_{50} \\
 = & -4.030E_{\text{HOMO}} + 0.977\text{ramp}(A \log P98 - 5.302) \\
 & + 60.173\text{ramp}(E_{\text{LUMO}} + 0.605) \\
 & - 0.573\text{ramp}(12.3146 - A \log P) - 27.035, \\
 & R^2 = 0.998; \text{ adjusted } R^2 = 0.998; \\
 & \text{cross validated } R^2 = 0.995; \\
 & \text{significant regression} = \text{yes}, \\
 & \text{critical SOR } F \text{ value} = 1089.
 \end{aligned}
 \tag{2}$$

In conclusion, the binding score for the tested compounds were congruent with their AChE inhibitory activity. A good correlation between the predicted and the experimentally observed inhibitory activities (pIC_{50}) (Tables 2 and 3) of the most ketoprofen-chalcone-amide hybrides suggested that the identified binding conformations of these inhibitors are reliable. The results of docking study provided an insight into the pharmacophoric structural requirements (compound **IX**) for AChE inhibitory activity of this class of compounds.

EXPERIMENTAL

Melting points are uncorrected and were measured on a Büchi melting point apparatus B-545 (Büchi-Labortechnik AG, Switzerland). NMR spectra were recorded on 400 (^1H) and on 150:91 MHz (^{13}C) spectrometers (Bruker, Germany) in DMSO- d_6 with TMS as internal standard and on the δ scale in ppm. Mass spectra (EI, 70 eV, and FAB) were recorded on MAT 8200 spectrometers (Finnegan MAT, USA). TLC plates 60 F254 were purchased from Merck. The chromatograms were visualized under UV 254–366 nm and iodine. For atom numbering refer to Fig. 2.

***N*-(4-Acetylphenyl)-2-(3-benzoyl)propanamide amide derivative of ketoprofen (VII).** A mixture of ketoprofen (**V**) (282 mg, 1.0 mmol) and PCl_5 (459 mg, 3.0 mmol) was stirred at 50°C, for 30 min, followed by addition of 4-amino acetophenone (270 mg, 2.0 mmol). The mixture was stirred at the same temperature for another 2 h. After cooling, few drops of water were added and stirred for 15 min, then the mixture was partitioned between CHCl_3 (3 \times 30 mL) and water (30 mL). The combined organic layers were dried (Na_2CO_3), filtered and evaporated to dryness. The residue was purified on a short SiO_2 column (5 g) using, in gradient, MeOH (0–10%) and CHCl_3 as eluent to give the desired amide (**VII**) (204 mg, 55%) as an amorphous.

General procedure for preparation of ketoprofen chalcone derivatives (VIII–XVII). To a stirred solution of ketone derivative (**VII**) (100 mg, 0.27 mmol) in EtOH (10 mL) was added arylaldehydes (0.37 mmol) and aq. solution of 2 M NaOH (5 mL). After stirring at ambient temperature for 24 h, the mixture was neutralized with 1 M HCl and partitioned with EtOAc (3–

Table 2. The observed and calculated IC_{50} of ketoprofen derivatives (**VIII**)–(**XVII**)

Compd.	pIC_{50} (obs.)	Pred. value	Residual
(VIII)	13.161	13.188	−0.07
(IX)	13.836	13.827	0.01
(X)	12.169	12.158	0.01
(XI)	11.315	11.320	0.00
(XII)	10.684	10.681	0.00
(XIII)	11.714	11.719	0.00
(XIV)	9.740	9.776	−0.04
(XV)	11.553	11.501	0.04
(XVI)	11.927	11.956	−0.02
(XVII)	10.758	10.722	0.04

15 mL). The combined organic extracts were washed with brine, dried over Na_2SO_4 and evaporated in vacuo. The residue was purified on a short SiO_2 column using the eluent hexane: EtOAc (3 : 2) as eluent to give the chalcone analogues (**VIII**–**XVII**).

2-(3-Benzoylphenyl)-*N*-(4-cinnamoylphenyl)propanamide (VIII). From benzaldehyde (39 mg). Yield: 102 mg (60%) as a yellow amorphous, $R_f = 0.64$, IR (film, ν , cm^{-1}): 3348 (NH), 1658 (C=O), 1597 (C=C); ^1H NMR, δ , ppm: 7.90–7.23 (m, 20H, Ar-H + $\text{H}_{\text{olefinic-7}}$ + $\text{H}_{\text{olefinic-8}}$), 3.31 (br s, 1H, H-18); 1.30 (s, 3H, Me-31); ^{13}C NMR, δ , ppm: 190.1 ($\text{C}_{24}=\text{O}$), 189.8 ($\text{C}_9=\text{O}$), 172.0 ($\text{C}_{17}=\text{O}$), 143.9 (C-7 + C-13), 136.5 (C-21), 134.8 (C-19 + C-25), 132.1, 131.2, 129.0, 128.4, 127.8, 127.1, 126.5, 126.0, 125.4, 124.0 (C_{arom}), 122.4 (C-8), 40.1 (C-18), 19.9 (Me-31); EIMS: m/z , 460 [$\text{M} + \text{H}$] $^+$.

2-(3-Benzoylphenyl)-*N*-(4-(3-(4-tolyl)acryloyl)phenyl)propanamide (IX). From 4-methylbenzaldehyde (44 mg). Yield: 114 mg (65%) as a brown solid, mp 66–68°C, $R_f = 0.69$, IR (film, ν , cm^{-1}): 2920 (CH), 3298 (NH), 1670 (C=O), 1600 (C=C); ^1H NMR, δ , ppm: 9.57 (s, 1H, NH), 8.51 (br s, 1H, $\text{H}_{\text{olefinic-7}}$), 8.10–7.26 (m, 18H, Ar-H + $\text{H}_{\text{olefinic-8}}$), 3.38 (br s, 1H, H-18), 2.03 (s, 3H, $\text{C}_1\text{-Me}$), 1.31 (s, 3H, Me-31); ^{13}C NMR, δ , ppm: 190.3 ($\text{C}_{24}=\text{O}$), 189.7 ($\text{C}_9=\text{O}$), 170.0 ($\text{C}_{17}=\text{O}$), 146.0 (C-7 + C-13), 137.9 (C-21), 135.9 (C-19 + C-25), 132.4, 132.0, 131.1, 129.5, 128.7, 128.5, 127.7, 126.0, 125.4 (C_{arom}), 124.2 (C-8), 41.9 (C-18), 26.4 (Me-1), 19.2 (C-31); EIMS: m/z , 474 [$\text{M} + \text{H}$] $^+$.

2-(3-Benzoylphenyl)-*N*-(4-(3-(4-methoxyphenyl)acryloyl)phenyl)propanamide (X). From 4-methoxybenzaldehyde (50 mg). Yield: 110 mg (61%), mp 120–122°C, $R_f = 0.70$, IR (film, ν , cm^{-1}): 3352 (NH), 2924 (CH), 1697 (C=O), 1593 (C=C); ^1H NMR, δ , ppm: 8.60 (br s, 1H, $\text{H}_{\text{olefinic-7}}$), 8.28 (br s, 2H, H-11 + H-15), 7.98–7.25 (m, 16H, Ar-H + $\text{H}_{\text{olefinic-8}}$), 3.68 (br s, 1H, H-18), 3.21 (s, 3H, OMe), 1.55 (s, 3H, Me-31);

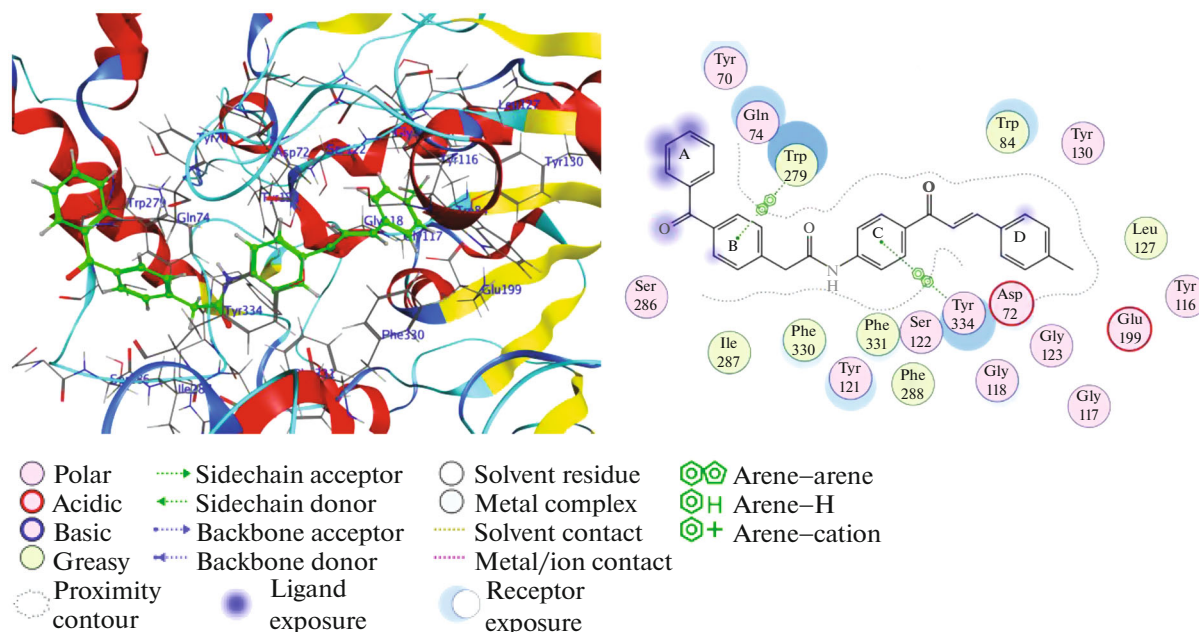


Fig. 3. Molecular modelling of compound (IX) with AChE generated by MOE.

^{13}C NMR, δ , ppm: 191.4 ($\text{C}_9=\text{O}$ + $\text{C}_{24}=\text{O}$), 171.7 ($\text{C}_{17}=\text{O}$), 153.6 ($\text{C}-\text{OMe}$), 145.8 ($\text{C}-7$ + $\text{C}-13$), 137.3 ($\text{C}-21$), 136.4 ($\text{C}-19$ + $\text{C}-25$), 132.4, 132.0, 129.4, 128.4, 127.3, 128.5, 127.7, 126.8, 125.8, 125.4 (C_{arom}), 124.6 ($\text{C}-8$), 115.2 ($\text{C}-2$ + $\text{C}_{\text{arom}}-6$), 53.1 (OMe), 42.8 ($\text{C}-18$), 19.9 ($\text{Me}-31$); EIMS: m/z , 490 [$\text{M} + \text{H}$] $^+$.

2-(3-Benzoylphenyl)-*N*-(4-(3-(4-hydroxyphenyl)acryloyl)phenyl)propanamide (XI). From 4-hydroxybenzaldehyde (45 mg). Yield: 98 mg (56%) as a brown solid, mp 107–109°C, R_f = 0.69, IR (film, ν , cm^{-1}): 3456 (OH), 3336, (NH), 1674 ($\text{C}=\text{O}$), 1597 ($\text{C}=\text{C}$); ^1H NMR, δ , ppm: 9.57 (s, 1H, OH), 8.54 (br s, 1H, $\text{H}_{\text{olefinic}}-7$), 7.98–7.28 (m, 16H, Ar–H + $\text{H}_{\text{olefinic}}-8$), 6.56 (br s, 2H, H-2 + H-6), 3.48 (br s, 1H, H-18), 1.37 (s, 3H, Me-31); ^{13}C NMR, δ , ppm: 190.9 ($\text{C}_{24}=\text{O}$), 189.7 ($\text{C}_9=\text{O}$), 171.8 ($\text{C}_{17}=\text{O}$), 161.4 ($\text{C}-\text{OH}$), 145.9 ($\text{C}-7$ + $\text{C}-13$), 137.4 ($\text{C}-21$), 136.5 ($\text{C}-19$ + $\text{C}-25$), 132.4, 132.1, 129.6, 129.4, 128.5, 127.5, 127.2, 126.9, 125.7, 125.5 (C_{arom}), 124.9 ($\text{C}-8$), 115.6 ($\text{C}-2$ + $\text{C}-6$), 43.0 ($\text{C}-18$), 19.5 ($\text{Me}-31$); EIMS: m/z , 476 [$\text{M} + \text{H}$] $^+$.

***N*-(4-(3-(4-Aminophenyl)acryloyl)phenyl)-2-(3-benzoylphenyl)propanamide (XII).** From 4-aminobenzaldehyde (45 mg). Yield: 117 mg (67%), mp 82–84°C, R_f = 0.67, IR (film, ν , cm^{-1}): 3376 (NH), 3335, 3228 (OH and NH_2), 3053 (CH), 1676 ($\text{C}=\text{O}$), 1589 ($\text{C}=\text{C}$); ^1H NMR, δ , ppm: 9.64 (br s, 2H, NH_2), 7.96 (br s, 1H, $\text{H}_{\text{olefinic}}-7$), 7.88–7.37 (m, 16H, H-11 + H-15 + Ar–H + $\text{H}_{\text{olefinic}}-8$), 6.64 (m, 2H, H-2 + H-6), 3.44 (br s, 1H, H-18), 1.38 (s, 3H, Me-31); ^{13}C NMR, δ , ppm: 191.2 ($\text{C}_9=\text{O}$ + $\text{C}_{24}=\text{O}$), 171.5 ($\text{C}_{17}=\text{O}$), 146.6 ($\text{C}-7$ + $\text{C}-13$ + $\text{C}-\text{NH}_2$), 137.4 ($\text{C}-21$), 136.6 ($\text{C}-19$ +

$\text{C}-25$), 132.2, 132.0, 129.8, 129.3, 128.3, 127.7, 127.5 (C_{arom}), 126.8 ($\text{C}-8$), 115.7 ($\text{C}-2$ + $\text{C}-6$), 43.2 ($\text{C}-18$), 19.8 ($\text{Me}-31$); EIMS: m/z , 475 [$\text{M} + \text{H}$] $^+$.

2-(3-Benzoylphenyl)-*N*-(4-(3-(4-dimethylamino)acryloyl)phenyl)propanamide (XIII). From 4-(*N,N*-dimethylamino)benzaldehyde (55 mg). Yield: 117 mg (63%), mp 110–112°C, R_f = 0.65, IR (film, ν , cm^{-1}): 3356 (NH), 2897 (CH), 1662 ($\text{C}=\text{O}$), 1597 ($\text{C}=\text{C}$); ^1H NMR, δ , ppm: 9.99 (s, 1H, NH), 7.97 (br s, 1H, $\text{H}_{\text{olefinic}}-7$), 7.91–7.38 (m, 16H, Ar–H + $\text{H}_{\text{olefinic}}-8$), 6.76 (m, 2H, H-2 + H-6), 3.48 (br s, 1H, H-18), 1.37 (s, 3H, Me-31); ^{13}C NMR, δ , ppm: 191.9 ($\text{C}_9=\text{O}$ + $\text{C}_{24}=\text{O}$), 171.7 ($\text{C}_{17}=\text{O}$), 146.3 ($\text{C}-\text{NMe}_2$), 144.9 ($\text{C}-7$ + $\text{C}-13$), 137.6 ($\text{C}-21$), 137.1 ($\text{C}-19$ + $\text{C}-21$ + $\text{C}-25$), 132.5, 132.1, 131.5, 130.0, 129.7, 128.4, 127.1, 127.2 (C_{arom}), 126.7 ($\text{C}-8$), 115.5 ($\text{C}-2$ + $\text{C}-6$), 43.8 ($\text{C}-18$), 19.9 ($\text{Me}-31$); EIMS: m/z , 503 [$\text{M} + \text{H}$] $^+$.

2-(3-Benzoylphenyl)-*N*-(4-(3-(4-nitrophenyl)acryloyl)phenyl)propanamide (XIV). From 4-nitrobenzaldehyde (56 mg). Yield: 136 mg (73%) as an orange-red solid, mp 186–188°C, R_f = 0.72, IR (film, ν , cm^{-1}): 3387 (NH), 2974 (CH), 1705 ($\text{C}=\text{O}$), 1597 ($\text{C}=\text{C}$); ^1H NMR, δ , ppm: 9.71 (s, 1H, NH), 8.47 (br s, 2H, H-2 + H-6), 8.32 (br s, 1H, $\text{H}_{\text{olefinic}}-7$), 7.89–7.32 (m, 16H, Ar–H + $\text{H}_{\text{olefinic}}-8$), 6.76 (m, 2H, H-3 + H-5), 3.46 (br s, 1H, H-18), 1.38 (s, 3H, Me-31); ^{13}C NMR, δ , ppm: 190.9 ($\text{C}_9=\text{O}$ + $\text{C}_{24}=\text{O}$), 171.9 ($\text{C}_{17}=\text{O}$), 147.8 ($\text{C}-\text{NO}_2$), 145.1 ($\text{C}-7$ + $\text{C}-13$), 137.6 ($\text{C}-21$), 137.0 ($\text{C}-19$ + $\text{C}-25$), 133.4, 132.5, 130.9, 129.8, 128.8, 127.4 (C_{arom}), 121.9 ($\text{C}-8$), 42.3 ($\text{C}-18$), 19.9 ($\text{Me}-31$); EIMS: m/z , 505 [$\text{M} + \text{H}$] $^+$.

2-(3-Benzoylphenyl)-N-(4-(3-(4-fluorophenyl)acryloyl)phenyl)propanamide (XV). From 4-fluorobenzaldehyde (46 mg). Yield: 115 mg (65%) as an oil, $R_f = 0.56$, IR (film, v, cm^{-1}): 3348 (NH), 2924 (CH), 1672 (C=O), 1597 (C=C), 829 (C–F); ^1H NMR, δ , ppm: 9.53 (s, 1H, NH), 8.51 (br s, 1H, $\text{H}_{\text{olefinic-7}}$), 7.98–7.26 (m, 18H, Ar–H + $\text{H}_{\text{olefinic-8}}$), 3.41 (br s, 1H, H-18), 1.31 (s, 3H, Me-31); ^{13}C NMR, δ , ppm: 191.2 ($\text{C}_9=\text{O}$ + $\text{C}_{24}=\text{O}$), 172.2 ($\text{C}_{17}=\text{O}$), 157.6 (d, $J_{\text{C,F}} = 247$ Hz, C–F), 144.8 (C-7 + C-13), 137.4 (C-19 + C-21 + C-25), 132.4, 132.0, 130.6, 129.5, 128.6, 128.4, 127.5, 126.7 (C_{arom}), 122.5 (C-8), 115.8 (C-2 + C-6), 43.0 (C-18), 19.9 (Me-31); EIMS: m/z , 477/479 [$\text{M} + \text{H}$] $^+$.

2-(3-Benzoylphenyl)-N-(4-(3-(4-chlorophenyl)acryloyl)phenyl)propanamide (XVI). From 4-chlorobenzaldehyde (52 mg). Yield: 120 mg (66%) as a yellow solid, mp 176–178°C, $R_f = 0.55$, IR (film, v, cm^{-1}): 3171 (NH), 1674 (C=O), 1593 (C=C), 833 (C–Cl); ^1H NMR, δ , ppm: 8.79 (s, 1H, NH), 8.36 (br s, 1H, $\text{H}_{\text{olefinic-7}}$), 8.07–7.32 (m, 18H, Ar–H + $\text{H}_{\text{olefinic-8}}$), 3.38 (br s, 1H, H-18), 1.37 (s, 3H, Me-31); ^{13}C NMR, δ , ppm: 191.5 ($\text{C}_{24}=\text{O}$), 189.4 ($\text{C}_9=\text{O}$), 171.9 ($\text{C}_{17}=\text{O}$), 144.9 (C-7 + C-13), 137.1 (C–Cl), 136.5 (C-19 + C-21 + C-25), 131.5, 130.6, 129.8, 128.5, 127.8, 126.5, 124.7 (C_{arom}), 122.3 (C-8), 43.8 (C-18), 19.7 (Me-31); EIMS: m/z , 494/496 [$\text{M} + \text{H}$] $^+$.

2-(3-Benzoylphenyl)-N-(4-(3-(4-bromophenyl)acryloyl)phenyl)propanamide (XVII). From 4-bromobenzaldehyde (69 mg). Yield: 122 mg (63%) as a pale yellow solid, mp 152–154°C, $R_f = 0.71$, IR (film, v, cm^{-1}): 3335 (NH), 3043 (CH), 1974 (C=O), 1585 (C=C), 594 (C–Br); ^1H NMR, δ , ppm: 9.86 (s, 1H, NH), 8.38 (br s, 1H, $\text{H}_{\text{olefinic-7}}$), 7.97–7.37 (m, 18H, Ar–H + $\text{H}_{\text{olefinic-8}}$), 3.36 (br s, 1H, H-18), 1.39 (s, 3H, Me-31); ^{13}C NMR, δ , ppm: 191.0 ($\text{C}_{24}=\text{O}$), 189.9 ($\text{C}_9=\text{O}$), 172.3 ($\text{C}_{17}=\text{O}$), 144.1 (C-7 + C-13), 139.0 (C-21), 133.3 (C-19 + C-25), 133.9, 132.4, 131.1, 130.5, 129.5, 129.3, 128.5, 127.5, 126.6, 124.8 (C_{arom}), 122.4 (C-8), 44.0 (C-18), 19.9 (Me-31); EIMS: m/z , 525/527 [$\text{M} + \text{H}$] $^+$.

AChE activity assay. Human serum AChE activity was determined using Ellman et al. [28] method as follows: 5,5'-dithiobis-(2-nitrobenzoic acid (DTNB) solution (50 μL , 0.001 M) was added to sodium phosphate buffer solution (2.25 mL, pH 7.3, 0.2 M), then serum (10 μL) was added, mixed well and 2 mL of the mixture was transferred to a measuring cell (1 cm). This procedure was followed by addition of acetyl thiocholine iodide (ASChI) (34 μL , 0.06 M). The changes in absorbency are measured before and after adding the substrate at (430 nm) for (3 min). The enzyme activity is calculated as concentration in μmole of the substrate hydrolyzed to each mL of samples in 3 min and expressed as $\mu\text{mole}/3 \text{ min}/\text{mL}$. The IC_{50} values were calculated by Bliss method [33] and expressed as mean \pm SD of the replicates.

CONCLUSIONS

We have synthesized a new series of ketoprofen-based chalcone derivatives derived ketoprofen as potential acetylcholinesterase (AChE) inhibitors. Compound 2-(3-benzoylphenyl)-N-(4-(3-(4-tolyl)acryloyl)phenyl) propanamide (**IX**) bearing methyl substituent at position C-4 of the phenyl group of the chalcone moiety showed significant inhibitory effect against the AChE. In general, substituents such as 4-methoxyphenyl, 4-hydroxyphenyl, 4-fluorophenyl, 4-fluorophenyl, and 4-chlorophenyl are beneficial for the activity of chalcone-amide derivatives derived ketoprofen. Introduction of 4-methylphenyl substituent in the chalcone group resulted in selective inhibitory effect against AChE. However, introduction of unsubstituted phenyl, 4-aminophenyl, 4-nitrophenyl and 4-bromophenyl moieties on chalcone scaffold does not improve the activity. Therefore, (**IX**) could be a promising lead as AChE inhibitor due to its potent activity against this enzyme.

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COMPLIANCE WITH ETHICAL STANDARDS

This article does not contain any studies involving human participants performed by any of the authors and does not contain any studies involving animals performed by any of the author.

Conflict of Interests

The authors declare that they have no conflicts of interest.

SUPPLEMENTARY INFORMATION

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REFERENCES

1. Thies, W. and Bleile, L., *Alzheimers Dement.*, 2013, vol. 9, pp. 208–245. <https://doi.org/10.1016/j.jalz.2013.02.003>
2. Prince, M., Ali, G.-C. Guerchet, M., Prina, M., Albanese, E., and Wu, T.-T., *Alzheimer's Res. Ther.*, 2016, vol. 8, pp. 23–35. <https://doi.org/10.1186/s13195-016-0188-8>
3. Hample, H., Mesuam, M.-M., Cuello, A.C., Farlow, M.R., Giacobini, E., Grossberg, G.T., Khachaturian, A.S., Vergallo, A., Cavado, E., Snyder, P.J., and Khachaturian, Z.S., *Brain*, 2018, vol. 141, pp. 1917–1933. <https://doi.org/10.1093/brain/awy132>

4. Anand, P., Singh, B., and Singh, N., *Bioorg. Med. Chem.*, 2012, vol. 20, pp. 1175–1180.
<https://doi.org/10.1016/j.bmc.2011.12.042>
5. Schuster, D., Spetea, M., Music, M., Rief, S., Fink, M., Kirchmair, J., Schutz, J., Wolber, G., Langer, T., Stuppner, H., Schmidhammer, H., and Rollinger, J.M., *Bioorg. Med. Chem.*, 2010, vol. 18, pp. 5071–5080.
<https://doi.org/10.1016/j.bmc.2010.05.071>
6. Coyle, J.T., Price, D.L., and De Long, M.R., *Science*, 1983, vol. 219, pp. 1184–1190.
<https://doi.org/10.1126/science.6338589>
7. Francis, P.T., Palmer, A.M., Snape, M., and Wilcock, G.K., *J. Neurol. Neurosurg. Psychiatry*, 1999, vol. 66, pp. 137–147.
<https://doi.org/10.1136/jnnp.66.2.137>
8. Lane, R.M., Potkin, S.G., and Enz, A., *Int. J. Neuropsychopharmacol.*, 2006, vol. 9, pp. 101–124.
<https://doi.org/10.1017/S1461145705005833>
9. Giacobini, E., *Pharmacol. Res.*, 2004, vol. 50, pp. 433–440.
<https://doi.org/10.1016/j.phrs.2003.11.017>
10. Cacabelos, R., Torrellas, C., Tejjido, O., and Carril, J.C., *Pharmacogenomics*, 2016, vol. 17, pp. 1041–1074.
<https://doi.org/10.2217/pgs-2016-0031>
11. Goyal, D., Kaur, A., and Goyal, B., *ChemMedChem*, 2018, vol. 13, pp. 1275–1299.
<https://doi.org/10.1002/cmdc.201800156>
12. Benek, O., Soukup, O., Pasdiorova, M., Hroch, L., Sepsova, V., Jost, P., Hrabanova, M., and Jun, D., *ChemMedChem*, 2016, vol. 11, pp. 1264–1269.
<https://doi.org/10.1002/cmdc.201500383>
13. Tumiatti, V., *J. Med. Chem.*, 2001, vol. 44, pp. 105–109.
<https://doi.org/10.1021/jm000991r>
14. Sangnoi, Y., Sakulkeo, O., Yuenyongsawad, S., Kanjana-opas, A., Ingkaninan, K., Plubrukarn, A., and Suwanborirux, K., *Mar. Drugs*, 2008, vol. 6, pp. 578–586.
<https://doi.org/10.3390/md20080029>
15. Liu, H.-R., Huang, X.-Q., Lou, D.-H., Liu, X.-J., Liu, W.-K., and Wang, Q.-A., *Bioorg. Med. Chem. Lett.*, 2014, vol. 24, pp. 4749–4753.
<https://doi.org/10.1016/j.bmcl.2014.07.087>
16. Zhao, F.-C., Wu, Y., and Song, X.-J., *Med. Sci. Monit.*, 2017, vol. 23, pp. 3311–3317.
<https://doi.org/10.12659/MSM.901842>
17. Díaz-Rubio, L., Hernández-Martínez, R., Estolano-Cobián, A., Chávez-Velasco, D., Salazar-Aranda, R., de Torres, N.W., Rivero, I.A., García-González, V., Ramos, M.A., and Córdova-Guerrero, I., *App. Sci.* 2019, vol. 9, pp. 410–439.
<https://doi.org/10.3390/app9030410>
18. Ismail, M.M., Kamel, M.M., Mohamed, L.W., Faggal, S.I., and Galal, M.A., *Molecules*, 2012, vol. 17, pp. 7217–7231.
<https://doi.org/10.3390/molecules17067217>
19. de Souza, G.A., da Silva, S.J., Del Cistia, C.N., Pitasse-Santos, P., Pires, L.O., Passos, Y.M., Cordeiro, Y., Cardoso, C.M., Castro, R.N., Sant’Anna, C.M.R., and Kümmerle, A.E., *J. Enzyme Inhib. Med. Chem.*, 2019, vol. 34, pp. 631–637.
<https://doi.org/10.1080/14756366.2019.1571270>
20. Zhou, X., Wang, X.-B., Wang, T., and Kon, L.-Y., *Bioorg. Med. Chem.*, 2008, vol. 16, pp. 8011–8021.
<https://doi.org/10.1016/j.bmc.2008.07.068>
21. Sonmez, F., Kurt, B.Z., Gazioglu, I., Basile, L., Dag, A., Cappello, V., Ginex, T., Kucukislamoglu, M., and Guccione, S., *J. Enzyme Inhib. Med. Chem.*, 2017, vol. 32, pp. 285–297.
<https://doi.org/10.1080/14756366.2016.1250753>
22. Guzior, N., Wi ckowska, A., Panek, D., and Malawska, B., *Curr. Med. Chem.*, 2013, vol. 22, pp. 373–404.
<https://doi.org/10.2174/0929867321666141106122628>
23. Anand, P. and Singh, B., *Arch. Pharm. Res.*, 2013, vol. 36, pp. 375–399.
<https://doi.org/10.1007/s12272-013-0036-3>
24. McHardy, S.F., Wang, H.-Y.L., McCowen, S.V., and Valdez, M.C., *Expert Opin. Ther. Pat.*, 2017, vol. 27, pp. 455–476.
<https://doi.org/10.1080/13543776.2017.1272571>
25. Sharma, K., *Mol. Med. Rep.*, 2019, vol. 20, pp. 1479–1487.
<https://doi.org/10.3892/mmr.2019.10374>
26. Zhuang, C., Zhang, W., Sheng, C., Zhang, W., Xing, C., and Miao, Z., *Chem. Rev.*, 2017, vol. 117, pp. 7762–7810.
<https://doi.org/10.1021/acs.chemrev.7b00020>
27. Willker, W., Leibfritz, D., Kerssebaum, R., and Birmel, W., *Mag. Res. Chem.*, 1993, vol. 31, pp. 287–292.
<https://doi.org/10.1002/mrc.1260310315>
28. Ellman, G.L., Courtney, K.D., Andresjr, V., and Featherstone, R.M., *Biochem. Pharmacol.*, 1961, vol. 7, pp. 88–90, IN1, 91–95.
[https://doi.org/10.1016/0006-2952\(61\)90145-9](https://doi.org/10.1016/0006-2952(61)90145-9)
29. *Molecular Operating Environment (MOE)*, ver. 2015.10, Chemical Computing Group Inc., Montreal, QC, Canada, 2015.
30. Barril, X. and Morley, S.D., *J. Med. Chem.*, 2005, vol. 48, 4432–4443.
<https://doi.org/10.1021/jm048972v>
31. *DMOL³ User Guide*, San Diego, CA, USA: Accelrys, Inc., 2003.
32. Rogers, D. and Hopfinger, A.J., *J. Chem. Inf. Comput. Sci.*, 1994, vol. 34, pp. 854–866.
<https://doi.org/10.1021/ci00020a020>
33. Bliss, C.I., *Microbiol. Mol. Biol. Rev.*, 1956, vol. 20, pp. 243–258.
<https://doi.org/10.1128/br.20.4.243-258.1956>